

Precision medicine: The future of diagnostic approach to pulmonary hypertension?

 Piotr Kedzierski,  Adam Torbicki

Department of Pulmonary Circulation, Thromboembolic Diseases and Cardiology, Centre of Postgraduate Medical Education, European Health Center-Otwock, Member of ERN-LUNG; Otwock-Poland

ABSTRACT

Pulmonary hypertension (PH) is a common finding that can result from many different pathological conditions. Depending on the etiology, treatment may be quite different, but early diagnosis and correct classification of PH is difficult. With an aging population and recently suggested decreased pulmonary arterial pressure threshold defining PH, we are facing even more diagnostic uncertainties. A new approach to patients' phenotyping is needed. Here we present available data and future perspectives on employing an in-depth analysis of the omics cascade to allow an earlier and more reliable diagnosis and classification of PH. Indeed, with the help of super-fast computing, it became possible to simultaneously consider the levels of thousands of potential biomarkers to find patterns specific for clinically suspected disease. The omics cascade is an invaluable source of information. However, while the genome can be perceived as providing possibilities, transcriptome—as carving them this is metabolome that may tell us “what is really going on” in an individual living organism. Metabolomics research requires blinded search for characteristic patterns of discrete changes in the levels of detectable metabolites. Since as many as 40,000 various substances are produced as a “side effect of staying alive”, metabolite profiling can be compared to fishing up for organized signals in a universe of chaos. Although difficult, such search for metabolic patterns that might lead to replacing the term *biomarker* by *metabolic fingerprinting* in the area of pulmonary circulation has already begun. (*Anatol J Cardiol* 2019; 22: 168-71)

Keywords: pulmonary hypertension, systems biology, metabolome, genome, transcriptome, proteome, epigenetics

Introduction

Despite a plethora of publications on candidate laboratory biomarkers, pulmonary hypertension (PH) cannot be diagnosed or properly classified by any single laboratory test. The DETECT trial is a good example of combining biomarkers including NT-proBNP in a stepwise approach to final diagnosis in a population at a moderately increased PAH risk (1). A relatively new concept, still in pre-clinical development, is based on systems biology, so-called *systemomics* (Fig. 1). This new holistic approach to living organisms is based on exploring their characteristics on several levels of organization. It has been recognized that although the genome determines hereditary or new mutation-dependent predispositions of an individual organism, it rarely fully determines the final phenotype (2). In pulmonary hypertension, such a strong influence of a single gene is exemplified by recently discovered EF12AK4 biallelic mutation, leading to the PVOD phenotype (3).

However, in most instances, the structural and functional properties of an individual organism are less directly related to genome being modified by epigenetic and environmental influences. As an example predisposition, to develop PH in sarcoidosis seems to be related to a signature involving 18 different genes (4).

The role of epigenetics in pulmonary hypertension has been studied mostly by the assessment of microRNA (miRNA, miRs), small non-coding RNA that downregulates the gene expression and in this way influences phenotypes. miRNA operates intracellularly, but it can be also detected and quantified in plasma. Some of the circulating miR were found to be decreased in PAH (150, 26a, 23 a, 125a) (5-8), while other (miR 130/301, and miR210) were increased in pulmonary circulation of PAH (9, 10). The miR patterns derived from different studies of PH are not always the same. A number of other downregulated circulating (miR-451, miR-1246) and upregulated cmiRNAs (miR-23b, miR-130a, miR-191) were identified in another study, suggesting that observed patterns may not have a universal clinical value (11). Indeed, relying

Address for correspondence: Adam Torbicki, MD, Department of Pulmonary Circulation, Thromboembolic Diseases and Cardiology, Centre of Postgraduate Medical Education, European Health Centre; Borowa Street 14/18, 05-400 Otwock-Poland
Phone: +48227103052 E-mail: adam.torbicki@ecz-otwock.pl

Accepted Date: 02.08.2019 **Available Online Date:** 25.09.2019

©Copyright 2019 by Turkish Society of Cardiology - Available online at www.anatoljcardiol.com
DOI:10.14744/AnatolJCardiol.2019.97820



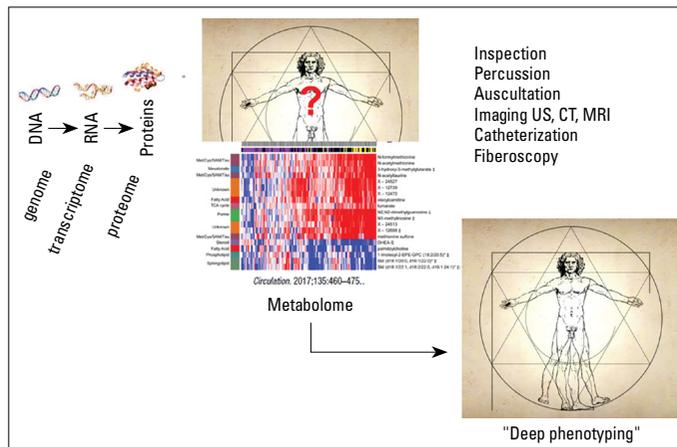


Figure 1. Systems biology - The future of medical diagnosis?

on plasma levels of cell-free miRs has been criticized because of the contamination risk (12). Therefore, although the role of miRs in the pathogenesis of PAH seems documented, some methodologic limitations together with the complexity of genetic and epigenetics interactions have not permitted the development of a clinically useful “epigenetic diagnostic test” for PH so far (13).

The proteome is another part of the systems biology. A recent trial used a machine-learning approach to analyze interrelations of 48 cytokines, chemokines, and growth factors using immunoassays (14). Patients in the PAH Group 1 (n=281) were divided into four clusters, considering the characteristic patterns of their proteomes without taking into account clinical features. A total of four PAH clusters with distinct proteomic immune profiles have been identified. Basic characteristics including age and gender, clinical PAH subtype, comorbidities, and pharmacological treatment were similar across clusters. One of the clusters had low cytokine levels similar to controls. Two other clusters were found to be related to high and low clinical risk category, respectively, as defined by classical prognostic stratification methods and assignment to a cluster predicted long-term outcomes. Those results were validated in an independent cohort of 104 patients from another institution. It is possible that precision medicine aimed at deep phenotyping of immune patchwork may help not only in risk stratification, but also in diagnosis and classification of patients with PH.

While the systems biology concept implies consideration of all levels of the omics cascade, this is the universe of metabolome, which seems to carry the most clinically promising information about the function and dysfunction of an individual organism. The genome can be perceived as providing possibilities, and the transcriptome as carving them; metabolome may tell us “what is really going on” in an individual living organism. Only part of this information is provided by “traditional phenotyping” relying on clinical observation, imaging, specific laboratory biomarker assessment, etc. Metabolome complements this information with “deep phenotyping”, and it could potentially reveal early and subtle problems, long before they can modify the traditionally assessed phenotype.

Metabolomics research requires blinded and not hypothesis-driven search for characteristic patterns of discrete modifications of levels of detectable metabolites. Since as many as 40,000 of various substances are produced as a side effect of “staying alive,” metabolite profiling can be compared to fishing up for organized signals in a universe of chaos. Search for metabolic patterns that might lead to replacing the term *biomarker* by metabolic *fingerprinting* is ready as a concept but still waiting for validation and implementation in the area of pulmonary circulation.

The existence of metabolomic heterogeneity of pulmonary arterial hypertension has been elegantly documented in tissue samples from the lungs explanted from PAH recipients when those were compared to the lung tissue sampled from patients with lung cancer. Metabolites were identified by means of liquid and gas chromatography and mass spectrometry. The lung tissue in patients with PAH presented disrupted glycolysis, an increased TCA (Krebs) cycle, and fatty acid metabolites with abnormal oxidation, suggesting that there are specific metabolic pathways likely contributing to the vascular remodeling process (15).

The lung tissue or intracellular matrix are not attractive sampling options for clinical medicine, but metabolome can also be assessed by sampling blood, urine, saliva, or even exhaled air. Drawing conclusions from plasma levels of metabolites seems more reasonable than doing so for microRNAs, which predominantly operate intra-cellularly. Several studies found distinct metabolic patterns in patients with PH using this approach. Lewis et al. (16) found that 21 out of 107 metabolites obtained from blood samples using of a multiplexed liquid chromatography mass spectrometry (LC-MS) system identified a characteristic pattern significantly associated with hemodynamic abnormalities assessed during RHC at rest and during exercise.

Bujak et al. (17) assessed metabolic fingerprints of 20 patients with PAH and compared them with matched healthy volunteers (n=20) using multiplatform metabolomics approach. Liquid chromatography and gas chromatography provided 21 and 9 metabolites, respectively, to form a PAH fingerprint. Some of those metabolites were related to energy imbalance, particularly glycolysis, but also to fatty acid, lipid, and amino acid metabolism. Interestingly, a profile consisting of 16 metabolites was confirmed as statistically significant in the validation study (17). Recently a much larger study found 53 circulating metabolites to distinguish patients with PAH (n=365) from healthy control subjects (n=121). Moreover, 20 out of 53 metabolites also discriminated patients with PAH from symptomatic patients in whom PH was excluded (n=139) (18). Sixty-two metabolites were related to the prognosis in PAH, majority acting independently from established prognostic markers. Also, in this study, metabolites related to bioenergetics were found to be of importance, which can be related to modified functional preferences of mitochondria in cells of the pulmonary arteries and right ventricle in patients with PAH (19).

The authors concluded that such deep phenotypic characterization of patients should be decisive for selecting and monitoring

treatment. Indeed, interventions correcting levels of some of the metabolites over time were associated with a better outcome (18).

Exhaled air can be transformed into a “breathprint” assessed with an “electronic nose”, and it has been used for the purpose of non-invasive analysis of metabolic derangements in PH. The part of metabolome that can be traced in breath is called “volatolome”, and it consists of volatile organic compounds (VOC). In a pilot case-control study, the volatolome not only differentiated 22 patients with PAH from 23 healthy volunteers, but it also identified those with idiopathic (n=15) and heritable (n=7, majority with the BMPR2 gene mutation) disease with an accuracy of 87%, suggesting a relationship between the breathprint and genetic mutations (20). So far, however, those preliminary observations have not been confirmed in any larger trial. The trial used the sensor-array approach to breath analysis, which is less informative than chromatography with mass spectrometry. Mansoor et al. (21) compared the composition of exhaled breath condensate of 27 idiopathic PAH patients with 30 healthy controls using mass spectrometry and created a model that discriminated between IPAH patients and controls with an accuracy of 75.4%.

Cikach et al. (22), using a modified mass spectroscopy method, found that among 21 analyzed organic compounds, 2-propanol, acetaldehyde, ethanol, pentane, 1-decene, 1-octene, and 2-nonene were either over- or underrepresented in patients with PAH compared with control subjects. The authors explored the exhaled volatolome of 31 patients with PAH and 34 controls applying mathematical discriminant analysis to VOC concentrations. The model was found to have an 86.1% accuracy for PAH detection. Ammonia received particular attention being overrepresented in PAH patients' exhaled breath compared to the control group, despite similar levels in blood. While interesting from the pathophysiological point of view, ammonia has been found overrepresented in other diseases as well, which limits its stand-alone diagnostic value as a VOC (22).

In a broader perspective, a metabolomics heterogeneity assessment could potentially provide a comprehensive (partly due to an automated approach to screening), early, and differential diagnosis of PAH. This could be particularly important when facing aging patient's population carrying multiple comorbidities of unclear contribution to increased pulmonary artery pressure commonly found in this clinical setting.

Characteristic patterns of metabolic modifications linked to deranged pathophysiological pathways operating in PAH would allow individualized treatment decisions based on deep phenotyping rather than on uncertain clinical and hemodynamic classification. Moreover, such treatment could consist of interventions ultimately modifying metabolome toward patterns compatible with health and good outcome.

However, many problems remain to be solved before systems biology will provide diagnostic tools for routine clinical practice in PH. While several teams found characteristic metabolic patterns in PAH, the “fingerprints” reported by different research teams are not identical, and sometimes, they are quite differ-

ent. The blame is placed on differences in analytical methods or environmental influences. Still, we are quite far from a validated, universally approved PAH metabolome. Furthermore, for obvious reasons, metabolomes were assessed in patients with unequivocal phenotypes, thus usually in late phases of PAH. Some of the modifications in metabolome profiles could be actually due to the nonspecific consequences of low cardiac output, hypoxemia, and venous stasis. Such a metabolome will be helpful neither for diagnosis nor differential diagnosis of PAH. Serial metabolic profiling of patients at risk for PAH (e.g., with a family history and/or carrying mutations, patients with scleroderma) could shed light on more specific patterns that can predict development of clinically unequivocal phenotypes. Such monitoring of the metabolome could provide also information useful “a rebours”, that is, in assessing the effects of treatment targeting pulmonary vascular disease. Finally, the presence or absence of specific PAH metabolome and its modifications during treatment would be of great value in patients with comorbidities to objectively assess whether specific treatment targeting pulmonary vascular disease is justified and effective.

Conclusion

A large number of candidate laboratory biomarkers derived from PH or PAH pathophysiology were tested for their clinical value. Individually, none was found useful for diagnostic purposes, except for the EIF2AK4 gene and NT-proBNP, the latter, however, as a part of stepwise diagnostic score. The future of laboratory biomarkers belongs to their contribution to “deep phenotyping” consisting of search for characteristic patterns in the genome, transcriptome, proteome, and/or metabolome of the patient (2, 23-26). Without neglecting the importance of genetic and epigenetic signatures, currently metabolomics emerges as the most informative area of systems biology research. Metabolome may provide a cross-reference confirming differential diagnosis made by standard phenotyping. In the future, metabolome may even provide information directly driving treatment decisions, while its modification over time may be considered as “deep monitoring” of treatment results.

So far, however, metabolic signatures have been assessed in well-defined, homogenous study populations. New research paradigms are now necessary to prove their value for early detection and differential diagnosis of PAH in real life.

Acknowledgment: Supported by scientific grant 501-1-54-01-12 of the Center of Postgraduate Medical Education.

Conflict of interest: None declared.

Peer-review: Internally peer-reviewed.

Authorship contributions: Concept – PK., A.T.; Design – PK., A.T.; Supervision – PK., A.T.; Fundings – None; Materials – None; Data collection

&/or processing – None; Analysis &/or interpretation – None; Literature search – P.K., A.T.; Writing – P.K., A.T.; Critical review – P.K., A.T.

References

1. Coghlan JG, Denton CP, Grünig E, Bonderman D, Distler O, Khanna D, et al. Evidence-based detection of pulmonary arterial hypertension in systemic sclerosis: the DETECT study. *Ann Rheum Dis* 2014; 73: 1340-9. [\[CrossRef\]](#)
2. Austin ED, West J, Loyd JE, Hemnes AR. Translational Advances in the Field of Pulmonary Hypertension Molecular Medicine of Pulmonary Arterial Hypertension. From Population Genetics to Precision Medicine and Gene Editing. *Am J Respir Crit Care Med* 2017; 195: 23-31. [\[CrossRef\]](#)
3. Eyries M, Montani D, Girerd B, Perret C, Leroy A, Lonjou C, et al. EIF2AK4 mutations cause pulmonary veno-occlusive disease, a recessive form of pulmonary hypertension. *Nat Genet* 2014; 46: 65-9.
4. Singla S, Zhou T, Javaid K, Abbasi T, Casanova N, Zhang W, et al. Expression profiling elucidates a molecular gene signature for pulmonary hypertension in sarcoidosis. *Pulm Circ* 2016; 6: 465-71.
5. Rhodes CJ, Wharton J, Boon RA, Roexe T, Tsang H, Wojciak-Stothard B, et al. Reduced microRNA-150 is associated with poor survival in pulmonary arterial hypertension. *Am J Respir Crit Care Med* 2013; 187: 294-302. [\[CrossRef\]](#)
6. Schlosser K, White RJ, Stewart DJ. miR-26a linked to pulmonary hypertension by global assessment of circulating extracellular microRNAs. *Am J Respir Crit Care Med* 2013; 188: 1472-5. [\[CrossRef\]](#)
7. Sarrion I, Milián L, Juan G, Ramon M, Furest I, Carda C, et al. Role of circulating miRNAs as biomarkers in idiopathic pulmonary arterial hypertension: possible relevance of miR-23a. *Oxid Med Cell Longev* 2015; 2015: 792846. [\[CrossRef\]](#)
8. Huber LC, Ulrich S, Leuenberger C, Gassmann M, Vogel J, von Blotzheim LG, et al. Featured Article: microRNA-125a in pulmonary hypertension: Regulator of a proliferative phenotype of endothelial cells. *Exp Biol Med* (Maywood) 2015; 240: 1580-9. [\[CrossRef\]](#)
9. Bertero T, Cottrill K, Krauszman A, Lu Y, Annis S, Hale A, et al. The microRNA-130/301 family controls vasoconstriction in pulmonary hypertension. *J Biol Chem* 2015; 290: 2069-85. [\[CrossRef\]](#)
10. White K, Lu Y, Annis S, Hale AE, Chau BN, Dahlman JE, et al. Genetic and hypoxic alterations of the microRNA-210-ISC1/2 axis promote iron-sulfur deficiency and pulmonary hypertension. *EMBO Mol Med* 2015; 7: 695-713. [\[CrossRef\]](#)
11. Wei C, Henderson H, Spradley C, Li L, Kim IK, Kumar S, et al. Circulating miRNAs as potential marker for pulmonary hypertension. *PLoS One* 2013; 8: e64396. [\[CrossRef\]](#)
12. Cheng HH, Yi HS, Kim Y, Kroh EM, Chien JW, Eaton KD, et al. Plasma processing conditions substantially influence circulating microRNA biomarker levels. *PLoS One* 2013; 8: e64795. [\[CrossRef\]](#)
13. Chun HJ, Bonnet S, Chan SY. Translational Advances in the Field of Pulmonary Hypertension. Translating MicroRNA Biology in Pulmonary Hypertension. It Will Take More Than "miR" Words. *Am J Respir Crit Care Med* 2017; 195: 167-78. [\[CrossRef\]](#)
14. Sweatt AJ, Hedlin HK, Balasubramanian V, Hsi A, Blum LK, Robinson WH, et al. Discovery of Distinct Immune Phenotypes Using Machine Learning in Pulmonary Arterial Hypertension. *Circ Res* 2019; 124: 904-19. [\[CrossRef\]](#)
15. Zhao Y, Peng J, Lu C, Hsin M, Mura M, Wu L, et al. Metabolomic heterogeneity of pulmonary arterial hypertension. *PLoS One* 2014; 9: e88727. [\[CrossRef\]](#)
16. Lewis GD, Ngo D, Hemnes AR, Farrell L, Doms C, Pappagianopoulos PP, et al. Metabolic Profiling of Right Ventricular-Pulmonary Vascular Function Reveals Circulating Biomarkers of Pulmonary Hypertension. *J Am Coll Cardiol* 2016; 67: 174-89. [\[CrossRef\]](#)
17. Bujak R, Mateo J, Blanco I, Izquierdo-Garcia JL, Dudzik D, Markuszewski MJ, et al. New Biochemical Insights into the Mechanisms of Pulmonary Arterial Hypertension in Humans. *PLoS One* 2016; 11: e0160505. [\[CrossRef\]](#)
18. Rhodes CJ, Ghataorhe P, Wharton J, Rue-Albrecht KC, Hadinnapola C, Watson G, et al. Plasma Metabolomics Implicates Modified Transfer RNAs and Altered Bioenergetics in the Outcomes of Pulmonary Arterial Hypertension. *Circulation* 2017; 135: 460-75. [\[CrossRef\]](#)
19. Culley MK, Chan SY. Mitochondrial metabolism in pulmonary hypertension: beyond mountains there are mountains. *J Clin Invest* 2018; 128: 3704-15. [\[CrossRef\]](#)
20. Cohen-Kaminsky S, Nakhleh M, Perros F, Montani D, Girerd B, Garcia G, et al. A proof of concept for the detection and classification of pulmonary arterial hypertension through breath analysis with a sensor array. *Am J Respir Crit Care Med* 2013; 188: 756-9. [\[CrossRef\]](#)
21. Mansoor JK, Schelegle ES, Davis CE, Walby WF, Zhao W, Aksenov AA, et al. Analysis of volatile compounds in exhaled breath condensate in patients with severe pulmonary arterial hypertension. *PLoS One* 2014; 9: e95331. [\[CrossRef\]](#)
22. Cikach FS Jr., Tonelli AR, Barnes J, Paschke K, Newman J, Grove D, et al. Breath analysis in pulmonary arterial hypertension. *Chest* 2014; 145: 551-8. [\[CrossRef\]](#)
23. Elinoff JM, Agarwal R, Barnett CF, Benza RL, Cuttica MJ, Gharib AM, et al. Challenges in Pulmonary Hypertension: Controversies in Treating the Tip of the Iceberg. A Joint National Institutes of Health Clinical Center and Pulmonary Hypertension Association Symposium Report. *Am J Respir Crit Care Med* 2018; 198: 166-74. [\[CrossRef\]](#)
24. Maron BA, Abman SH. Translational Advances in the Field of Pulmonary Hypertension. Focusing on Developmental Origins and Disease Inception for the Prevention of Pulmonary Hypertension. *Am J Respir Crit Care Med* 2017; 195: 292-301.
25. Nakhleh MK, Haick H, Humbert M, Cohen-Kaminsky S. Volatolomics of breath as an emerging frontier in pulmonary arterial hypertension. *Eur Respir J* 2017; 49: pii: 1601897. [\[CrossRef\]](#)
26. Newman JH, Rich S, Abman SH, Alexander JH, Barnard J, Beck GJ, et al. Enhancing Insights into Pulmonary Vascular Disease through a Precision Medicine Approach. A Joint NHLBI-Cardiovascular Medical Research and Education Fund Workshop Report. *Am J Respir Crit Care Med* 2017; 195: 1661-70. [\[CrossRef\]](#)